Potential Aroma in Several Varieties of Spanish Grapes

Elvira López-Tamames,^{*,†} Nieves Carro-Mariño,[‡] Yusuf Ziya Gunata,[§] Claude Sapis,[§] Raymond Baumes,[§] and Claude Bayonove[§]

Institut National de la Recherche Agronomique, Institut des Produits de la Vigne, Laboratoire des Arômes et des Substances Naturelles, 9 Place Viala, F-34060 Montpellier, France, Unitat de Nutrició i Bromatologia, CERTA, Facultat de Farmàcia, Universitat de Barcelona, Avenida Joan XXIII s/n, E-08028 Barcelona, Spain, and Departamento de Química Analítica, Nutrición y Bromatología, Facultad de Química, Universidad de Santiago de Compostela, Avenida de las Ciencias s/n, E-15706 Santiago de Compostela, Spain

Free and bound aroma compounds of eight Spanish grape cultivars (Albariño, Treixadura, Listan, Viura, Xarel.lo, Parellada, Garnacha, and Tempranillo) used in winemaking were studied. With a single-extraction procedure it was possible to provide information about the potential flavor of these varieties: bound aroma was determined directly by TFA glycoside analysis and indirectly by enzyme hydrolysis release of aglycon. An aroma data base of these varieties and a comparison between the grape flavor composition of each variety were established. Grape characteristics (sugars, acidity, polyphenols, juice yield), mainly due to climate and cultivar factors, were related with the aroma composition. Non-terpenyl compounds such as 2-phenylethanol and benzyl alcohol were the main glycosidically bound compounds of Spanish varieties. Direct determination of glycosides is better than hydrolysis of aglycon as an approach to the potential aroma of grapes.

Keywords: Aroma; glycosides; Spanish grape cultivars; Albariño; 'Treixadura; Listan; Viura; Xarel.lo; Parellada; Garnacha; Tempranillo

INTRODUCTION

Quality wines have different flavor properties which depend on varietal characteristics. This typical flavor is mainly due to volatile compounds that proceed from the grape and whether they are in free volatile form (Williams *et al.*, 1981) or in bound form, usually as glycoside (Williams *et al.*, 1982a,b; Voirin *et al.*, 1992a). Bound aroma, potentially developed during winemaking, is unknown for several grape cultivars.

French varieties such as Muscat (Carro-Mariño et al., 1995), Chardonnay (Sefton et al., 1993; Arrhenius et al., 1996), and Cabernet Sauvignon (Gómez et al., 1995) and German varieties such as Riesling (Skouroumounis & Winterhalter, 1994) have been the cultivars most studied in the establishment of data bases of flavor compounds; however, the majority of Italian and Spanish varieties are unknown. Only few data exist about free aroma of winemaking products of Spanish cultivars such as Viura, Xarel.lo, Parellada (De la Presa et al., 1995) and Albariño (Versini et al., 1995). In the current study, the free and bound aroma of eight typical Spanish grapes (Albariño, Treixadura, Listan, Viura, Xarel.lo, Parellada, Garnacha, and Tempranillo) was screened to learn their varietal aroma. These grape varieties have world areas of cultivation estimated at <5 000, <5 000, 28 000, 60 000, 43 000, <5.000, 378 000, and 85 000 ha, respectively. Although some of them have small areas of cultivation, they are used in the production of wines of recognized high quality (with Appellation of Origin Controlled, AOC) and are known worldwide. The grapes considered in this study are from two different climate zones: Galician (northwestern Spain), with an Atlantic climate (high humidity and rain), and Mediterranean, with a climate with high insolation and temperature.

Several studies (Gunata et al., 1985; Strauss et al., 1987; Voirin et al., 1992a; Sefton et al., 1993; Williams et al., 1995; Zhou et al. 1996) have reported methods of analyzing wine flavor. However, few references describe a single and routine method which allows a quantification of the free and bound aroma of grapes, mainly non-Muscat varieties. In this research, the extraction and separation were carried out according to the method proposed by Gunata et al. (1985) and aroma compounds were analyzed using the same grape sample. Later, free compounds of grape extracts were analyzed by GC and MS. Bound compounds of grape extracts were determined with two procedures: a direct method, which determined glycosides by trifluoroacetylation (TFA), based on Voirin (1992b) but modified to eliminate the interferences of two glycoside isomers (Carro-Mariño et al., 1995), and an indirect method, which determined the aglycons released before enzyme hydrolysis. Moreover, since climate and cultivar factors affect aroma composition (De la Presa et al., 1995; Gómez et al., 1995; García-Jarés et al., 1995; Arrhenius et al., 1996), these variables were also considered by including more than one sample of the same variety: a data base of grape aromas of Spanish varieties was reported taking into account the variation due to harvest and geography.

MATERIALS AND METHODS

Plant Material. Eight Spanish grape varieties (Albariño, Treixadura, Listán, Viura, Xarel.lo, Parellada, Garnacha, and Tempranillo) were considered. Albariño, Treixadura, and Listán grapes were harvested in 1993 from different Galician AOC cultivars: *Rias Baixas AOC* [Albariño (A) grapes came from three zones: Valle del Salnés (As), Rosal (Ar), and Condado del Tea (Ac)] and *Ribeiro AOC* [Listán] (L) and Treixadura (Tr)]. Galician varieties came from a region with a Atlantic climate with an high humidity and rain, while the

^{*} Author to whom correspondence should be addressed (e-mail elopez@farmacia.far.ub.es).

[†] Universitat de Barcelona.

[‡] Universidad de Santiago de Compostela.

[§] Institut National de la Recherche Agronomique.

sample ^a	variety (color)	origin	AOC (origin controlled)	sugars (g/L)	soluble solids (° Brix)	titratable acidity (g of H ₂ SO ₄ /L)	pН	maturity index (° Brix/ titratable acidity)	poly- phenols (280 nm)	juice yield (mL of juice/kg of grape)
Ac	Albariño (white)	Galician	Rias Baixas	197	22.0	3 50	3 68	6.3	30.3	0 755
Ar	Albariño (white)	Galician	Rias Baixas	175	18.5	4.45	3.42	4.2	24.3	0.785
As	Albariño (white)	Galician	Rias Baixas	174	18.4	6.70	3.28	2.7	19.9	0.630
Gp	Grenache (red)	Mediterranean	Penedès	244	24.5	2.50	3.47	9.8	28.7	0.740
Gĥ	Grenache (red)	Mediterranean		153	16.5	6.05	3.15	2.7	17.2	0.640
L	Listan (white)	Galician	Ribeiro	145	15.8	2.30	3.57	6.9	24.8	0.710
P92	Parellada (white)	Mediterranean	Penedès & Cava	156	16.8	3.40	3.21	4.9	10.4	0.760
P93	Parellada (white)	Mediterranean	Penedès & Cava	146	15.9	2.30	3.49	6.9	12.3	0.810
T92	Tempranillo (red)	Mediterranean	Penedès	212	21.8	2.25	3.98	9.7	29.0	0.720
T93	Tempranillo (red)	Mediterranean	Penedès	170	18.1	2.60	3.67	7.0	24.8	0.780
Tr	Treixadura (white)	Galician	Ribeiro	128	14.3	4.35	3.58	3.3	22.2	0.845
V92	Viura (white)	Mediterranean	Penedès & Cava	149	16.2	1.95	3.43	8.3	10.4	0.770
V93	Viura (white)	Mediterranean	Penedès & Cava	163	17.4	2.05	3.70	8.5	15.1	0.820
X92	Xarel.lo (white)	Mediterranean	Penedès & Cava	148	16.1	3.15	3.33	5.1	11.6	0.790
X93	Xarel.lo (white)	Mediterranean	Penedès & Cava	175	18.5	2.25	3.62	8.2	12.1	0.835

^a c, r, s, p, h: geographical origin. c: Condado del Tea. r: Rosal. s: valle del Salnés. p: Penedès. h: Hérault. 92, 93: year of harvest.

μ_{μ}

		identification															
	Kovats index	a, retention		Galici		Mediterranean varieties											
	(Carbowax phase)	time b, MS	Ac	Ar	As	L	Tr	Gp	Gh	P92	P93	T92	T93	V92	V93	X92	X93
1 hexanal	1127	a,b	57	1032	741	199	34	416	844	257	201	294	564	139	44	479	317
2 (<i>E</i>)-hex-3-en-1-al	1148	a.b	2	14	16	1	1	3	2	3	3	4	5	10	3	3	2
3 (Z)-hex-2-en-1-al	1236	a.b	8	40	24	12	8	208	9	122	11	110	14	109	13	189	23
4 (E)-hex-2-en-1-al	1243	a.b	349	2373	1300	587	193	401	1624	436	1124	371	941	231	215	715	686
5 hexanol	1365	a.b	400	843	485	151	449	346	119	194	407	441	263	83	136	207	160
6 (E)-hex-3-en-1-ol	1376	a,b	8	15	13	3	6	3	7	5	10	5	6	1	1	5	4
7 (Z)-hex-3-en-1-ol	1395	a.b	26	81	81	128	35	59	21	73	125	70	78	91	69	50	10
8 (E)-hex-2-en-1-ol	1410	b	799	1153	809	352	415	162	52	318	686	273	313	65	132	327	207
9 (Z)-hex-2-en-1-ol	1416	a.b	5	4	19	3	5	9	4	9	7	1	5	3	1	5	5
10 <i>trans</i> -furanic linalool oxid	1449	a.b		1			1	1	1		1	1					
11 <i>cis</i> -furanic linalool oxid	1476	a.b		_		1	-	2	1	1	-	3					
12 benzaldehvde	1538	a.b	4	13	9	8	3	16	10	6	4	5	3	15	2	17	4
13 linalool	1558	a.b	1	2	3	5		1	2	2	1	1					
14 hotrienol	1680	b.	2	1	1	-	1	1	1	1	1	1	1	1		1	
15 α-terpineol	1697	a.b	4	1	1	1	3	Ô	2	-	1	2	1	-	2	-	1
16 7-ethoxy-3,7-dimethyl-1,5- octadien-3-ol	1725	b	1	4	2	2	1	1	2	1	1	ĩ	1	1	~	1	1
17 <i>trans</i> -pyranic linalool oxid	1755	b	7	13	17	3	1	1	1	1		1			2		2
18 cis-pyranic linalool oxid	1772	b		1	1			1	1	2	1	1	2	1	3	1	
19 citronellol	1777	a.b	1	3	2	2	1	6	2		2	2	1	4		4	1
20 nerol	1809	a.b	1	2	2	1		0	11	2		2		4	1	3	2
21 hexanoic acid	1857	b.	206	199	131	26	69	138	14	144	91	43	40	88	67	135	97
22 geraniol	1858	a.b	13	24	16	11	18	14	17	10	3	6	4		2	15	5
23 benzyl alcohol	1877	a.b	30	59	66	19	32	66	44	111	28	26	16	49	23	167	50
24 2-phenylethanol	1910	a.b	64	76	79	92	27	56	63	84	22	50	17	49	13	87	27
25 2,6-dimethyl-3,7-octadiene- 2.6-diol	1958	a,b	3	16	15	6	3	49	10	64	13	17	9	11	1	7	6
26 2.6-dimethyl-7-octen-2.6-diol	2027	a.b	3	8	4	4	3	5	4	11	9	7	7	3	3	13	2
27 2,6-dimethyl-1,7-octadiene- 3,6-diol	2219	b	2	2	1	2	2		29		5	1	2		1		3
28 2,6-dimethyl-7-octene- 1,6-diol	2266	b	1	2	1	1	1	1	1	2	2		1				1
29 (E)-2,6-dimethyl-2,7- octadiene-1,6-diol	2298	b	2	5	2	3	3	7	13	5	23	14	6	10	6	10	15
30 (Z)-2,6-dimethyl-2,7- octadiene-1,6-diol	2343	b	11	17	10	9	8	7	3	9	5	3	2	3	1	6	7
31 geranic acid	2567	b	1	1		1		12	60	19	3	10		2		8	1
32 $\tilde{3}$ -hydroxy- β -damascone	2613	b	3	3	3	4	2	10	13	4	3	4	2		3		4
33 unknown	2620	а	1		2			1	9			2		7		5	1
34 vanillin	2625	b	3	9	4	6	2	26	8	33	7	9	4	28	4	22	46
35 3-oxo-α-ionol	2637	b	2	3	1	1	2	5	1	2	1	7	1	2	1	9	

^a A: Albariño. L: Listán. Tr: Treixadura. G: Garnacha. P: Parellada. T: Tempranillo. V: Viura. X: Xarel.lo. 92, 93: year of harvest. c, r, s, p, h: geographical origin.

other varieties were collected from an area with a Mediterranean climate. The Garnacha grape variety (G) was harvested in 1992 from two areas: domaine du Chapître ENSA-INRA, Herault (Gh), France, and vineyards in the *Penedès AOC* (Gp). Viura (V), Xarel.lo (X), Parellada (P), and Tempranillo (T) grapes also came from *Penedès AOC* and were harvested in 1992 and 1993 (V92, V93, X92, X93, P92, P93, T92, and T93, respectively). Albariño, Treixadura, Listán, Viura, Xarel.lo, and Parellada are white varieties, and Garnacha and Tempranillo are red varieties. All grape samples were collected on the cellar before winemaking. They were frosted rapidly with liquid nitrogen and stored at -20 °C until crushing.

Juice Preparation. Grape berry samples (1 kg) were

Table 3. Bound Compounds (Released by Enzyme Hydrolysis and Glycoside Form) in Spanish Varieties^a (in $\mu g/L$)

	Galician varieties					Mediterranean varieites										
	Ac	Ar	As	L	Tr	Gp	Gh	P92	P93	T92	T93	V92	V93	X92	X93	
aglycons released by hydrolysis																
36 <i>trans</i> -furanic linalool oxid	1	4	2				2		2		1		3		2	
37 cis-furanic linalool oxid	1	1	1						1		1					
38 linalool	2	10	9	2	1		2	5	6		5	13	13	4	16	
39 hotrienol		1	1			1	1	2	1			1	1	1	1	
40 α-terpineol		1	2				2		1		1		1	1	1	
41 <i>trans</i> -pyranic linalool oxid	1	5	2						1							
42 citronellol	1	2	1				2		1		1		1		1	
43 nerol	1	3	2				2	1	1		2	1	2	1	3	
44 geraniol	8	29	21	3	5	2	23	8	4	2	4	3	5	7	10	
45 benzyl alcohol	16	65	58	18	18	79	193	193	67	44	24	181	108	283	129	
46 2-phenylethanol	24	36	37	40	11	22	69	52	31	21	16	70	52	61	43	
47 2,6-dimethyl-3,7-octadiene-2,6-diol	3	13	5	1			1	4	1			1	1	3	1	
48 2,6-dimethyl-7-octen-2,6-diol	1	2	1	1		1	1		3	1	1	3	4	1	2	
49 2,6-dimethyl-1,7-octadiene-3,6-diol		1	1						1							
50 2,6-dimethyl-7-octene-1,6-diol	1	3	1	1		2	3	5	1	2		11	1	7	2	
51 (E)-2.6-dimethyl-2.7-octadiene-1.6-diol	5	6	3	3	1	1	7	6	1	2	1	5	2	7	3	
52 (Z)-2.6-dimethyl-2.7-octadiene-1.6-diol	32	38	19	23	10	3	11	8	3	3	3	7	8	17	13	
53 geranic acid	3	8	3	1	1	2	5	14	2	5	4	2	9	4		
54 3-hydroxy- β -damascone		1	1			2	1	19	1	2		4		10		
glycosides																
55 benzvl β -D-glucopyranoside	500	681	289	1419	2462	652	578	1015	1512	554	439	316	923	709	1802	
56 linalyl β -D-glucopyranoside	22	30	16	25	6	10			7	5		7	15	7	6	
57 2.6-dimethyl-1.7-octadiene-3.6-diol β -D-	15									5			12	5	4	
glucopyranoside																
58 2-phenylethyl β -p-glucopyranoside	448	185	130	831	180	125	281	97	134	123	67	129	82	34	291	
59 nervl β -D-glucopyranoside		34	9									9	15		9	
60 geranyl β -D-glucopyranoside	139	28	5	93	26	14			11		11	11	19		9	
61 citronellyl β -D-glucopyranoside	6	18	4	29	9	2			12		4	11	19		-	
62 α -terpinevl β -D-glucopyranoside	53	22		22	16	20	10	7	6	6		14	11		13	
63 (R)-linalyl rutinoside																
64 benzyl rutinoside	332	99	32	140	139	245	183	187	455	152	137	232	413	64	288	
65 (S)-linalyl rutinoside	17		11	45	42	64	62	36	55	26	23	24	130	8	114	
66 benzyl 6- O -(α -L-arabinofuranosyl) β -D-	190	26	30	147	246	633	431	155	119	89	35	178	358	124	517	
g ucopyranoside + nerv rutinoside																
67 2-phenylethyl rutinoside	226	2810	439	718	571	651	2989	94	2915	247	672	805	1366	229	3108	
68 nervl 6- O -(α -L-arabinofuranosvl) β -D-	298	91	8	27	17	25	36	• •	260	13	0.2	22	36	9	52	
glucopyranoside + α -terpinevl rutinoside																
69.2 -phenylethyl $6-Q-(\alpha-L-arabinofuranosyl)$	735	1802	180	326	401	163	469	127	1537	203	162	316	1290	130	818	
β -p-glucopyranoside																
70 geranyl 6- O -(α -L-arabinofuranosyl)	211	21	20	25	12		55	13	35	9		6	13	37	65	
β -D-glucopyranoside	~	~1	~0	20	-~			10		5		5	10			
71 α -terpinevi 6-O-(α -L-arabinofuranosvi)	119	3	10	64	86	43	59	10	40	22		47	253	35	140	
β -D-glucopyranoside	-	-	-			-		-	-			-			-	

^a See Table 2 footnotes.

defrosted (4 °C overnight) and crushed. Cloudy juices were centrifuged (10 min at 7500*g*) at the same temperature and then filtered. Clear juice volumes were measured to calculate the juice yield (Table 1), expressed as mL of juice/kg of grape. Moreover, juice characteristics [sugar content (g/L), soluble solids (°Brix), titratable acidity (g of H₂SO₄/L), maturity index (°Brix/titratable acidity), pH, and total polyphenols (absorbance at 280 nm)] were established.

Extraction of Free and Bound Flavor. 200 mL of must [to which 10 μ L of 4-nonanol (Fluka 98%) solution (3.6 mg/ mL of methanol) was added as internal standard] was eluted through an Amberlite XAD-2 column (120 mm \times 7 mm i.d.) with 1.5 mL/min as flow rate. Free compounds were collected by elution with 50 mL of pentane/dichloromethane (2:1, v/v), and the bound fraction was obtained by elution with 50 mL of ethyl acetate. The free fraction was dried over sodium sulfate and then concentrated to 500 μ L by Dufton columns. Extracts were stored at -20 °C until analysis by GC. The bound fraction was diluted with ethyl acetate at 100.0 mL, and then two 50.0 mL portions were obtained. All extractions were performed in triplicate.

Enzyme Hydrolyses. One 50.0 mL portion of the ethyl acetate fraction was dried over sodium sulfate and evaporated at 30 °C under vaccum, then redissolved in 100 μ L of 0.1 M citrate-phosphate buffer (pH 5.0) and washed five times with 1 mL of pentane/dichloromethane (2:1, v/v) to remove traces of free compounds. 100 μ L of Pektolase 3PA (Grindsted) solution was added [1.2 mg of Pektolase 3PA in 100 μ L of 0.1

M citrate-phosphate buffer (pH 5.0)], and the mixture was incubated at 40 °C for 16 h. After incubation, released aglycons were extracted four times with 200 μ L of pentane/ dichloromethane (2:1, v/v) and 10 μ L of 4-nonanol solution (3.6 mg/mL of methanol) was added as internal standard. Aglycons were dried over sodium sulfate and then concentrated to 500 μ L by Dufton columns. Extracts were stored at -20 °C until analysis by GC.

Glycoside Determination. The other 50.0 mL portion of the ethyl acetate fraction was dried over sodium sulfate and evaporated at 30 °C under vaccum and then redissolved in 2.0 mL of bidistillated water and treated with 150 mg of polyvinylpolypyrrolidone (PVPP) (Polyclar AT, GAF) to remove probable polyphenol interferences; this mixture was magnetically stirred for 1 h at 4 °C with, and then PVPP was eliminated by cellulose filtration. The extract was dried on a Réacti-vial of 2 mL capacity at 60 °C with a nitrogen stream. Trifluoroacetylation was carried out at 60 °C (20 min) using 20 μ L of anhydrous pyridine and 20 μ L of MBTFA reagent [*N*-methylbis(trifluoroacetamide)] (Sigma). Just before GC analysis, 10 μ L of phenyl β -D-glucopyranoside (1 g/L of methanol) (Sigma 98%) was added as internal standard.

Chromatographic Method of Free Volatiles and Aglycons. A Varian model 6000 provided with a CP-Wax 52 CB, fused-silica capillary column (Chrompack) (25 m \times 0.3 mm i.d.; 1.2 μ m bonded phase) was used to perform the on-column GC analysis: injector temperature was programmed from 20 to 250 °C with a rate of 180 °C/min; column temperature was



Figure 1. Exponential regression between the content of bound geraniol and the maturation index. *Upper and lower limits of 95% confidence interval for the three analytical replicates were considered for the samples values.

programmed at 2 °C/min from 60 °C (3 min isothermal) to 220 °C and at 3 °C/min from 220 °C to 245 °C; hydrogen at 2.5 mL/min was used as carrier gas; FID temperature was 250 °C. Electron impact mass spectra were obtained by coupling the CP-Wax 52 CB, fused-silica capillary column to a Finnigan MAT ITD 700. The transfer line, heated at 240 °C, consisted of an open-split GC-ITD interface at atmospheric pressure and a flow restrictor which was a DB-5 fused-silica capillary column (1.15 m \times 0.15 mm i.d.; bonded phase). The temperature program used was the same as for the GC analysis. The

source temperature was 220 °C. Mass spectra were scanned between 50 and 80 eV in the range m/z 41–250 at 2-s intervals. Identification was also performed by comparison with Kovats Index of standards (Table 2).

Chromatographic Method of Glycosides. Equipment used was a Varian model 3300 with DB 5 fused-silica capillary column (30 m \times 0.32 mm) and an on-column injector. Injector temperature was programmed at 60 °C/min from 90 to 280 °C. Column temperature was programmed from 125 to 220 °C at 3 °C/min and from 220 to 280 °C at 2 °C/min with hydrogen as carrier gas at 1.4 mL/min. Flame ionization detector temperature was 300 °C. Identification was carried out by GC coupling to MS (Hewlett-Packard 5989 MS Engine) with the chromatographic conditions described previously but with helium (1 mL/min) as carrier gas. Source temperature was 250 °C, mass spectra were scanned at 70 eV in the range m/z 29–850 at 2-s intervals and by retention times comparison of standards (Voirin, 1992a).

Quantitation. Concentrations of free fraction (Table 2) and aglycones enzymatically hydrolyzed from glycosides (Table 3) were calculated using 4-nonanol as internal standard and they were expressed by average of three repeated analytical assays in μ g/L. Glycosides (Table 3) were quantified by phenyl β -D-glucopyranoside as internal standard, taking into account the extraction efficiency calculated with a reference solution fixed on Amberlite resin. The average of three repeated analytical assays was considered.

Statistical Analysis. Principal component analysis (PCA) was carried out using STATGRAPHICS 7.0. PCA considers as quantitative variables 71 aroma compounds, expressed in μ g/L. In order to consider the relationship between aroma composition and ripening, sugars, titratable acidity, polyphenols, and juice yield were also included as a quantitative parameters. Qualitative variables were variety (A, G, L, Tr, P, T, V, and X), year of harvest (92 and 93), and geographical origin in Spain (Galician and Mediterranean). Moreover, multiple correlation was performed to establish the relationship between the free and bound aroma components related to grape characteristics.



Component 1 (37.4%)

Figure 2. PCA of herbaceous compounds in some Spanish varieties.



Figure 3. PCA of free aroma compounds in some Spanish varieties.



	10 trans-furanic linalool oxid
	11 cis-furanic linalool oxid
	12 benzaldehyde
	13 linalool
	14 hotrienol
	15 α -terpineol
	16 7-ethoxy-3,7-dimethyl-1,5-octadien-3-ol
Albariño Condai Albariño Rosal	17 trans-pyranic linalool oxid
Albariño Salnés	18 cis-pyranic linalool oxid
Listan	19 citronellol
Garnacha Penedès	20 nerol
Garnacha Herault	21 hexanoic acid
Parellada 92 Parellada 93	22 geraniol
Tempranillo 92	23 benzyl alcohol
Tempranillo 93 Viura 92	24 2-phenylethanol
Viura 93	25 2,6-dimethyl-3,7-octadiene-2,6-diol
Xarel.lo 92	26 2,6-dimethyl-7-octen-2,6-diol
Xarei.io 93	27 2,6-dimethyl-1,7-octadiene-3,6-diol
	28 2,6-dimethyl-7-octene-1,6-diol
	29 (E)-2,6-dimethyl-2,7-octadiene-1,6-diol
	30 (Z)-2,6-dimethyl-2,7-octadiene-1,6-diol
	31 geranic acid
	32 3-hydroxy-β-damascone
	33 unknown
	34 vanilline

- 35 3-oxo- α -ionol
- Garnacha Penedès Garnacha Herault Parellada 92 Parellada 93 Tempranillo 92 Tempranillo 93

Albariño Condal

- Viura 93

- Xarel.lo 92
- 36 trans-furanic linalool oxid
- 37 cis-furanic linalool oxid

- 41 trans-pyranic linalool oxid
- 45 benzyl alcohol
- 46 2-phenylethanol
- 47 2,6-dimethyl-3,7-octadiene-2,6-diol
- 48 2,6-dimethyl-7-octen-2,6-diol
- 49 2,6-dimethyl-1,7-octadiene-3,6-diol
- 50 2,6-dimethyl-7-octene-1,6-diol
- 51 (E)-2,6-dimethyl-2,7-octadiene-1,6-diol
- 52 (Z)-2,6-dimethyl-2,7-octadiene-1,6-diol
- 53 geranic acid
- 54 3-hydroxi-β-damascone

Figure 4. PCA of the bound compounds released by enzymic hydrolysis in some Spanish varieties.





Figure 5. PCA of glycosides in some Spanish varieties.

RESULTS AND DISCUSSION

Table 2 shows the free aroma of Spanish grapes, expressed by mean $(\mu g/L)$, which correspond to the three analytical replicates. The bound aroma (released by enzyme hydrolysis and in glycoside form) of the Spanish grapes is expressed also by mean $(\mu g/L)$ and displayed in Table 3. Penedès grape varieties have a higher (p < p0.05) maturation index and more juice yield than Galician cultivars (mainly Albariño variety). The different ripening stages (Table 1) among varieties and within the same variety affect the level of several aroma compounds: for example, in Figure 1 the exponential regression between maturation index and the concentration of geraniol released by enzymic hydrolysis shows that the greater the ripening of the grape, the lower the amount of bound geraniol.

Free Aroma Fraction. Non-terpenyl compounds were the most abundant substances in the aroma-free fraction of Spanish varieties (Table 2): alcohols and aldehydes of six carbon atoms (benzyl alcohol, 2-phenylethanol, benzaldehyde, and hexanoic acid). Geraniol and several diols were the terpenes that appeared in higher concentration. Norisoprenoids such as 3-hydroxy- β -damascone and 3- α -oxoionol are present in several cultivars. This free aroma composition indicates that these cultivars have a nonfloral aroma profile with respect to the Muscat cultivars.

Alcohols and aldehydes of six carbon atoms (Figure 2) have a greasy odor, and their origin is mainly in the lipoxygenase activity of the grape (Cordonnier, 1989). PCA shows that these compounds were correlated (p <0.05). Hexanal (p < 0.05), (*E*)-hex-2-en-1-al (p < 0.04), and (E)-hex-3-en-1-ol (p < 0.01) were herbaceous compounds that seem related to the maturation index of the grape: the lower the acidity, the lower the level of these substances. The concentration of greasy alcohols and

	Albariño Condal
Ц	Albariño Rosal
	Albariño Salnés
<u>*</u>	Listan
ō	Garnacha Penedès
Õ	Garnacha Herault
*	Parellada 92
☆	Parellada 93
	Tempranillo 92
Ň.	Viura 92
ŏ	Viura 93
Δ	Xarel.lo 92
Δ	Xarel.lo 93
55	benzyl β-D-glucopyranoside
56	linalyl β -D-glucopyranoside
57	2,6-dimethyl-1,7-octadiene-3,6-diol β -D-glucopyranoside
58	2-phenylethyl β-D-glucopyranoside
59	neryl β -D-glucopyranoside
60	geranyl β-D-glucopyranoside
61	citronellyl β-D-glucopyranoside
62	α -terpineyl β -D-glucopyranoside
63	(R) linaly! rutinoside
64	benzyl rutinoside
65	(S) linalyl rutinoside
66	benzyl 6-O-(α -L-arabinofuranosyl)- β -D-glucopyranoside + neryl rutinoside
67	2-phenylethyl rutinoside
68	neryl 6-O-(α -L-arabinofuranosyl)- β -D-glucopyranoside + α -terpineyl rutinoside
69	2-phenylethyl 6-O-(α -L-arabinofuranosyl)- β -D-glucopyranoside
70	geranyl 6-0-(α -L-arabinofuranosyl)- β -D-glucopyranoside
71	α -terpinevi 6-O-(α -L-arabinofuranosvi)- β -D-glucopyranoside

П

aldehydes depends on the grape cultivar (p < 0.01): Albariño had the greatest levels, although there were variations due to the geographical origin or to the ripening stage; Listan, Xarel.lo, and Viura have the smallest quantities of greasy compounds. No differences in these alcohols and aldehydes were found between white and red cultivars.

Terpenes [linalool with their trans-furanic linalool oxide (p < 0.001)] and non-terpenyl substances (benzyl alcohol, 2-phenylethanol, benzaldehyde, norisoprenoids, geranic acid) were also correlated (p < 0.05). Galician cultivars, mainly Albariño, had more terpenyl compounds (*trans*-pyranic linalool oxid and several diols) than the Mediterranean varieties (Figure 3). Free aroma composition depended on grape cultivar: Listan had more linalool, Garnacha had more 3-hydroxy- β damascone, Xarel.lo had more vanillin, and Parellada had more (E)-2,6-dimethyl-2,7-octadiene-1,6-diol. Furthermore, the year of harvest changed free aroma composition: 1993 grapes had fewer aroma substances than 1992 grapes; this could be explained by the different maturation index of the grapes between years: terpenyl compounds, mainly geraniol, were less when acidity was lower (p < 0.005) (Figure 1).

Bound Aroma Fraction. Quantitatively, bound aroma released by enzyme hydrolysis was lower than the direct glycoside composition (Table 3). A possible explanation of this observation could be a partial hydrolysis of the enzyme. The aglycons released in the highest level are non-terpene: benzyl alcohol and 2-phenylethanol. Moreover, glycosides of benzyl alcohol and 2-phenylethanol were the highest glycosides. Voirin (1992b) indicates that the varieties richest on this nonterpene glycosides are neutral cultivars. PCA of bound aroma compounds released by enzyme hydrolysis (Figure 4) shows two groups of bound substances: terpenyl,

linalool oxide, and diol aglycones and other aglycon groups with benzyl alcohol, 2-phenylethanol, and geranic acid. Quantitatively, differences in the released aglycons were found when the variety (Albariño was richest in bound terpenyl aroma) and the year of harvest were considered. However, only the content of bound 2-phenylethanol and α -terpineol seems related to the maturation index: relationships between these compounds and their acidity (r = 0.8137 and r = 0.9017, respectively) were found for the grapes of the same Albariño variety.

PCA of grape glycosides is displayed in Figure 5. Two groups of glycosides are shown: glucopyranosides (except the benzyl β -D-glucopiranoside) and arabinosides with rutinosides; multiple correlations between both groups were found (p < 0.05). Furthermore, glycoside composition was highly variable and depended on the geographical origin and/or the year; for example, Viura, Xarel.lo, and Parellada harvested in 1993 have arabinoside and rutinoside values higher than those found in 1992; this difference could be explained by the degree of ripening of the grapes.

CONCLUSIONS

Free and potential aroma of eight Spanish varieties of grapes were established using a single extraction. The potential aroma was determined by two procedures: direct analysis of glycosides and indirect analysis of aglycons released by hydrolysis; indirect analysis was not representative of aroma precursors. Non-terpenyl compounds were the most abundant aroma substances in the considered Spanish grapes. Moreover, aroma components were quantitatively affected by the maturation index. Differences due to varietal and climate characteristics were shown; Galician grapes, mainly Albariño, were richest in varietal aroma.

ACKNOWLEDGMENT

We are grateful to Galician and Penedès wineries (Segura Viudas S.A.) for their collaboration.

LITERATURE CITED

- Arrhenius, S. P.; Mc Closkey, L. P.; Sylvan, M. Chemical markers for aroma of *Vitis vinifera* var. Chardonnay regional wines. J. Agric. Food Chem. **1996**, 44 (4), 1085–1090.
- Carro-Mariño, N.; López-Tamames, E.; García-Jarés, M. C. Contribution to the study of the aromatic potential of three Muscat Vitis vinifera varieties: Identification of new compounds. Food Sci. Technol. Int. 1995, 1, 105–116.
- Cordonnier, R. Mécanismes et facteurs de formation des composés à flaveurs herbacées. *Rev. Oenol.* **1989**, *53S*, 25–27.
- García-Jarés, C. M.; García-Martín, M. S.; Carro-Mariño, N.; Cela-Torrijos R. GC–MS identification of volatile components of Galician (Northwestern Spain) white wines. Application to differenciate Rías Baixas wines from wines produced in nearby geographical regions. *J. Sci. Food Agric.* **1995**, 69 (2), 175–184.
- Gómez, E.; Martínez, A.; Laencina, J. Changes in volatile compounds during maturation of some grape varieties. J. Sci. Food Agric. 1995, 67 (2), 229–233.

- Gunata, Y. Z.; Bayonove, C. L.; Baumes, R. L.; Cordonnier, R. E. I. The aroma of grapes. I. Extraction and determination of free and glycosidically bound fractions of some grape aroma components. J. Chromatogr. 1985, 331, 83–90.
- Presa de la, C.; Lamuela-Raventós, R. M.; Buxaderas, S.; Torre-Boronat, M. C. Differentiation and grouping characteristics of varietal grape musts from Penedès region (I). *Am. J. Enol. Vitic.* **1995**, *46* (3), 283–291.
- Sefton, M. A.; Francis, I. L.; Williams, P. J. The volatile composition of Chardonnay juices: A study by flavor precursor analysis. *Am. J. Enol. Vitic.* **1993**, *44* (4), 359–370.
- Skouroumounis, G. K.; Winterhalter, P. Glycosidically bound norisoprenoids from *Vitis vinifera* cv. Riesling. *J. Agric. Food Chem.* **1994**, *42* (5), 1068–1072.
- Statgraphics 7.0, Statistical graphics systems, Magnugisties Inc.: Rockville, MD, 1993.
- Strauss, C. R.; Gooley, P. R.; Wilson, B.; Williams, P. J. Application of droplet countercurrent chromatography to the analysis of conjugated forms of terpenoids, phenols, and other constituents of grape juice. *J. Agric. Food Chem.* **1987**, 35, 519–524.
- Versini, G., Orriols, I.; Serra-Adalla, D. Aroma components of Galician Albariño, Loureira and Godello wines. *Vitis* 1995, 33 (3), 165–170.
- Voirin, S. V.; Baumes, R.; Gunata, Y.; Bitteur, S.; Bayonove, C. Analytical methods for monoterpene glycosides in grape and wine. I. XAD-2 extraction and gas chromatographic– mass spectrometric determination of synthetic glycosides. J. Chromatogr. 1992a, 590, 313–328.
- Voirin, S. G.; Baumes, R. L.; Sapis, J. C.; Bayonove, C. Analytical methods for monoterpene glycosides in grape and wine. II. Qualitative and quantitative determination of monoterpene glycosides in grape. *J. Chromatogr.* 1992b, 595, 269–281.
- Williams, P. J.; Strauss, C. R.; Wilson, B. Classification of the monoterpenoid composition of Muscat grapes. Am. J. Enol. Vitic. 1981, 32, 3.
- Williams, P. J.; Strauss, C. R.; Wilson, B.; Massy-Westropp, R. A. Novel monoterpene disaccharide glycosides of *Vitis vinifera* grapes and wines. *Phytochemistry* **1982a**, *21*, 2013– 2020.
- Williams, P. J.; Strauss, C. R.; Wilson, B. Use of C18 reversedphase liquid chromatography for the isolation of monoterpene glycosides and norisoprenoid precursors from grape juice and wine. J. Chromatogr. 1982b, 235, 471–480.
- Williams, P. J.; Cynkar, W.; Francis, I. L.; Gray, J. D.; Iland, P. G.; Coombe, B.G. Quantification of glycosides in grapes, juices, and wines through a determination of glycosil glucose. J. Agric. Food Chem. **1995**, 43 (1), 121–128.
- Zhou, Y. W.; Riesen, R.; Gilpin, C. S. Comparison of Amberlite XAD-2/Freon-11 extraction with liquid–liquid extraction for the determination of wine flavor components. J. Agric. Food Chem. 1996, 44 (3), 818–82.

Received August 5, 1996. Accepted January 28, 1997.^{\otimes} This study was made possible by financial assistance from the Ministerio de Educación y Ciencia (Spain) and from the COMETT program for funding E.L-T. and N.C.M., respectively, during their stays in France.

JF960572W

[®] Abstract published in *Advance ACS Abstracts*, March 15, 1997.